



**Wings of Worry:
Avian Influenza Viruses**

Daniel R. Perez, PhD
Poultry Diagnostic and Research Center,
Department of Population Health,
College of Veterinary Medicine,
University of Georgia

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Acknowledgements

Flu lab (present)	USDA - ARS	U. Cambridge, UK
<i>L. Okhai, A. Baker, T. M. Miles, D. K. Regmi, A. Mark, I. Scaranotti, D. Schofield, L. He, K. Chrzastek, A. Ospina Jimenez</i>	<i>Amy Baker, David Suarez, Darrell Kapczynski, Adolfo Garcia-Sastre, Florian Krammer, John Hopkins U., Sabra Klein et al.</i>	<i>David R. Burke, Eugene Skepner, R. Medina, D. Steinhauer, L. Byrd-Leotis, Emory U., K. Koelle, S. Mark Tompkins, D. E. Stallknecht, J. C. Paulson, A. J. Thompson, R. Cummings, T. Mckinlay, L. J. Swayne, M. Molinari, Virginia Tech, L. Marr, N. Shetty, St. Jude CRH, R. Webb, NCBII-NIH, M. Nelson</i>
Flu lab (past members)	U. Rochester	U. London, UK
<i>F. Cargnin Faccin, D. Rajao, R. Palomares, L. Joshi, E. Howerth, S. Compton, ABSL-3 Personnel - AHRC, Jazmin, Hannah, Karly, Trey, and Morgan.</i>	<i>A. Sant et al.</i>	<i>R. G. Webster, P. Digard, U. Glasgow, UK, U. Edinburgh, UK, N. Lewis, I. Brown, P. Murcia, R. Fouchier, U. del Valle, Guatemala, C. Cordon-Rosales, UNC, Argentina, J. Echenique, P. T. M. Domingo, A. Perez, J. Cappuccio, A. Rimondi, U. Sao Paulo, Brazil, E. Arruda et al.</i>
Flu lab (past members)	CDC	CEIIRR
<i>F. Cargnin Faccin, D. Rajao, R. Palomares, L. Joshi, E. Howerth, S. Compton, ABSL-3 Personnel - AHRC, Jazmin, Hannah, Karly, Trey, and Morgan.</i>	<i>J. Stevens, H. Davies et al.</i>	<i>C. Caswell Eidson endowment funds, Perez Lab</i>

2

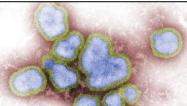
**Wings of Worry:
Avian Influenza Viruses**

- Bird Flu
- Zoonotic transmission of avian origin FLUAVs
- Modified live attenuated vaccines (MLVs) for mass vaccination strategies against avian influenza, including H5N1 HPAIV
- Aerosol exposure system – focus on MLV delivery
- Discussion of Jersey cows experimentally infected with H5N1 HPAIV
- Conclusions and final thoughts

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International Committee on Taxonomy of Viruses

Realm: Riboviria
Kingdom: Orthornavirae
Phylum: Negeriviricota
Subphylum: Polyiviricota
Class: Inflaviricetes
Order: Articulavirales



Digital-colorized negative-stained transmission electron micrograph of avian influenza viruses. Credit: CDC/R.A. Murphy

- Lipid bilayer envelope derived from host plasma membrane
- Pleomorphic particles with a diameter in the range 80–120 nm
- RNA genome, segmented, negative sense, RNA-dependent RNA polymerase complex

Family: Orthomyxoviridae

Genus: *Alphainfluenzavirus* – type A - FLUAV

Genus: *Betainfluenzavirus* – type B - FLUBV

Genus: *Deltafluenzavirus* – type D - FLUDV

Genus: *Gammainfluenzavirus* – type C - FLUCV

Genus: *Isavirus*

Genus: *Mykissvirus*

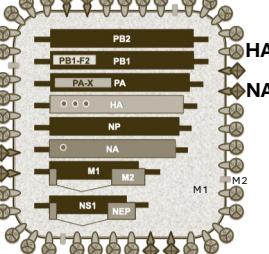
Genus: *Quaranjevirus*

Genus: *Sardinivirus*

Genus: *Thogotovirus*

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Influenza viruses are constantly changing



- Hemagglutinin (HA) and Neuraminidase (NA)**
- Surface glycoproteins
- Divided into subtypes
- 17 HA subtypes (avian)
- 9 NA subtypes (avian)
- 153 theoretical combinations
- H17, H18, H9-like, N10, N11, N2-like (fruit bats)
- Error Prone Polymerase**
- Point Mutations – Changes on surface proteins (HA) leads to Antigenic drift and Evasion of immune responses
- Segmented Genome = Reassortment**
- Reassortment leads to *Antigenic Shift* = Exchange of HA subtype
1918 H1N1, 1957 H2N2, 1968 H3N2, 1977 H1N1, 2009 pH1N1

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Flu Can Spread Between Animals (and People!)

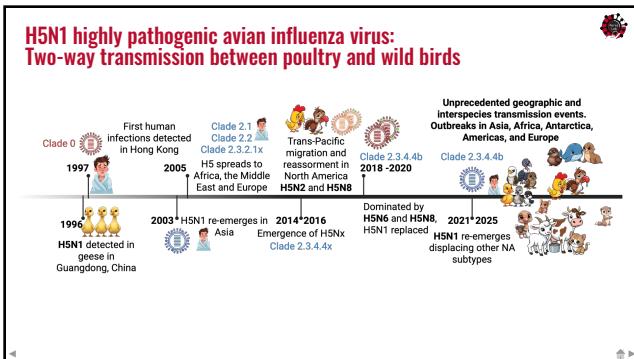
<img alt="Diagram showing the spread of influenza viruses between animals and people. It highlights the Hemagglutinin (HA) protein and its subtypes (H1-H10, H13, H14, H17, H18, H20, H21, H22, H23, H24, H25, H26, H27, H28, H29, H30, H31, H32, H33, H34, H35, H36, H37, H38, H39, H40, H41, H42, H43, H44, H45, H46, H47, H48, H49, H50, H51, H52, H53, H54, H55, H56, H57, H58, H59, H60, H61, H62, H63, H64, H65, H66, H67, H68, H69, H70, H71, H72, H73, H74, H75, H76, H77, H78, H79, H80, H81, H82, H83, H84, H85, H86, H87, H88, H89, H90, H91, H92, H93, H94, H95, H96, H97, H98, H99, H100, H101, H102, H103, H104, H105, H106, H107, H108, H109, H110, H111, H112, H113, H114, H115, H116, H117, H118, H119, H120, H121, H122, H123, H124, H125, H126, H127, H128, H129, H130, H131, H132, H133, H134, H135, H136, H137, H138, H139, H140, H141, H142, H143, H144, H145, H146, H147, H148, H149, H150, H151, H152, H153, H154, H155, H156, H157, H158, H159, H160, H161, H162, H163, H164, H165, H166, H167, H168, H169, H170, H171, H172, H173, H174, H175, H176, H177, H178, H179, H180, H181, H182, H183, H184, 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Avian Influenza (Bird Flu): A Threat to Birds and The Economy

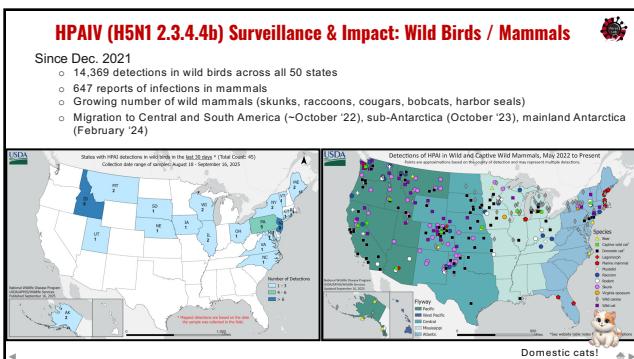
- Originally termed fowl plague in 1878
- Results in significant economic losses due to disease/deaths and culling to control infection
- Isolates are classified based on pathogenicity in domestic chickens (terminology adopted in 1981)
- Low pathogenic avian influenza (LPAI) viruses:**
 - Most isolates are low pathogenic – H9N2 LPAI viruses are the most prominent
 - Associated with outbreaks of varying intensity in domestic birds
- Highly pathogenic avian influenza (HPAI) viruses:**
 - Circulation of LPAI viruses that evolve into HPAI viruses
 - Restricted to H5 and H7 subtypes
 - HA with polybasic cleavage site
 - Intravenous Pathogenicity Index test of ≥ 1.2 (75% mortality)

Daniel M. Perez et al., (2018); Avian Influenza Virus

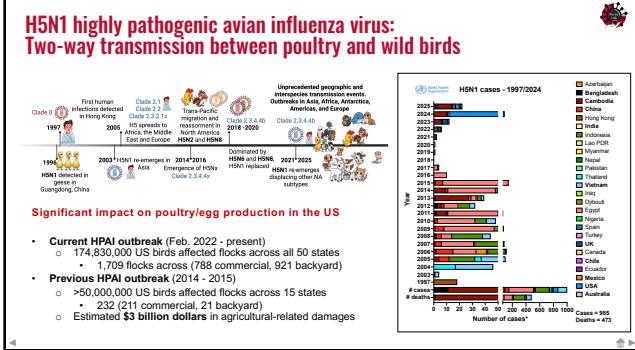
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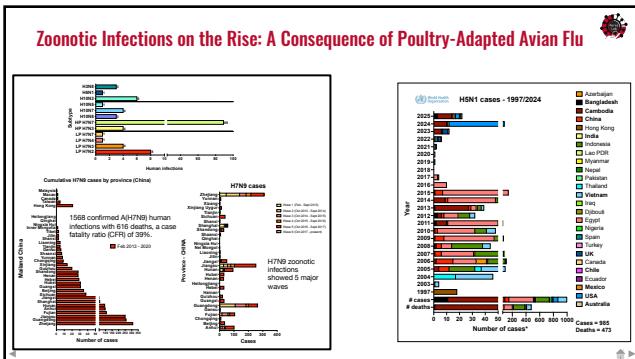
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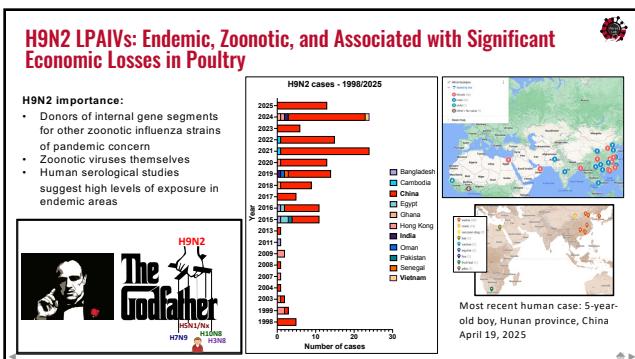
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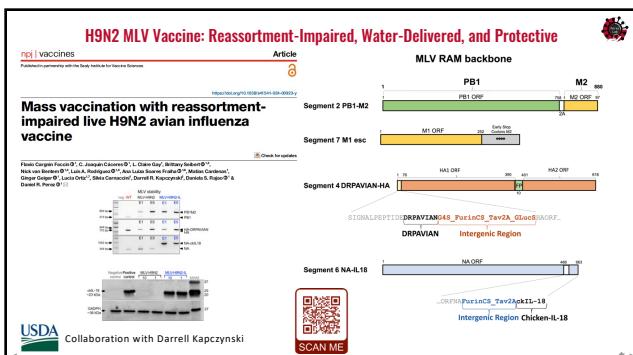
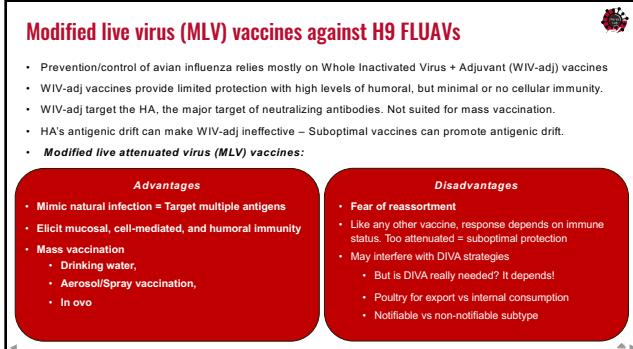
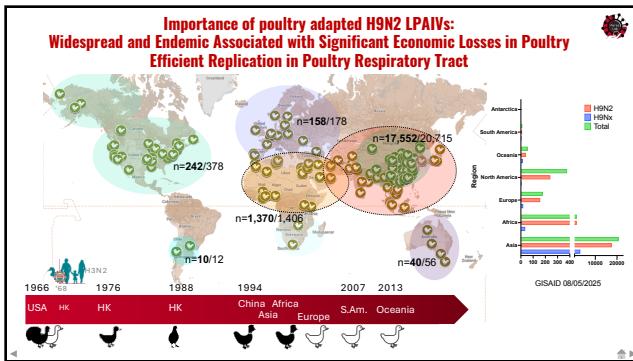
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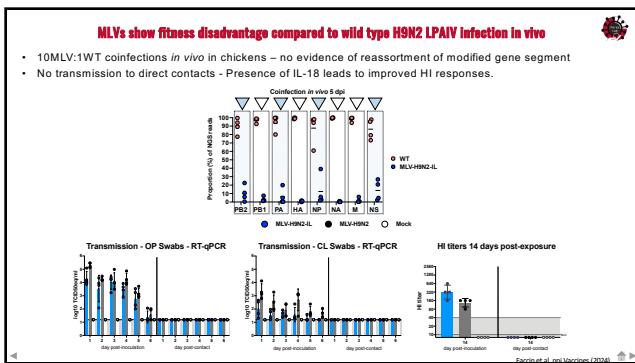


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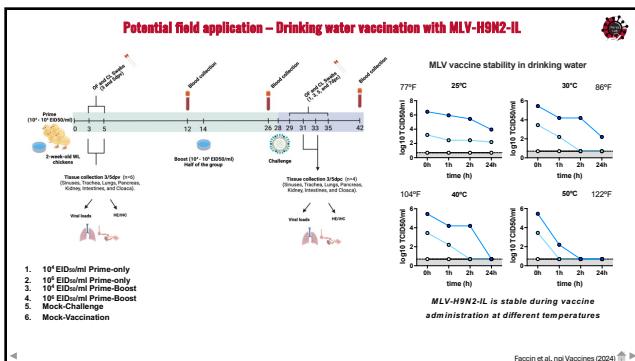


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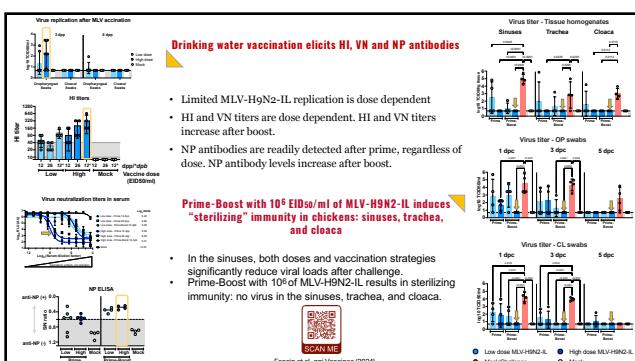




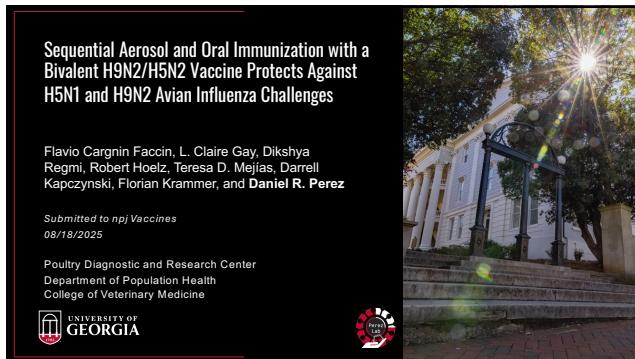
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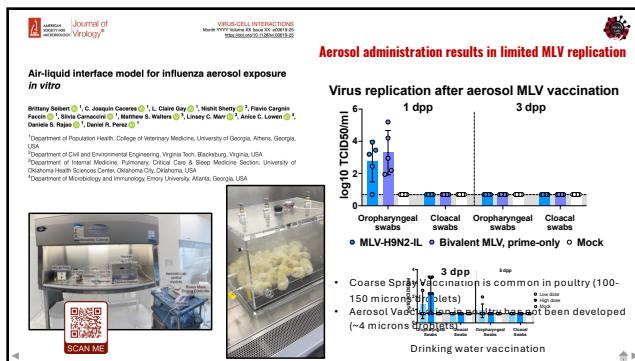
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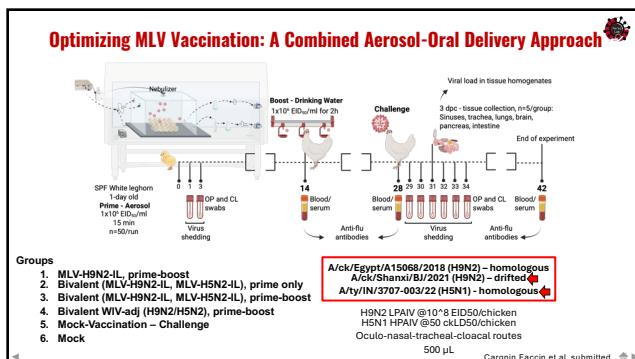
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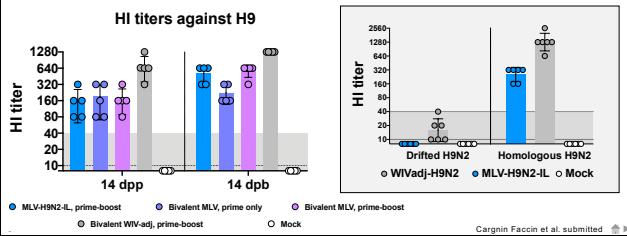
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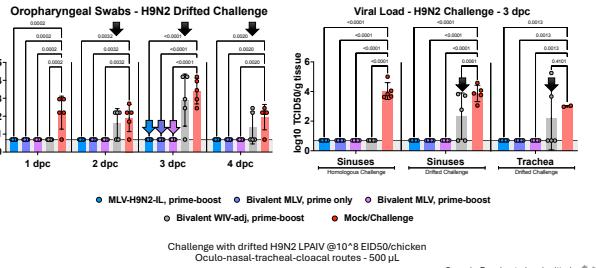
Aerosol/drinking water vaccination induced discernible HI responses against the homologous but not antigenically drifted H9 HA

- Prime vaccination via aerosol induced HI titers against H9
- Boost via drinking water increased antibody titers against H9
- Antibody titers against antigenically drifted strain ≤ 40



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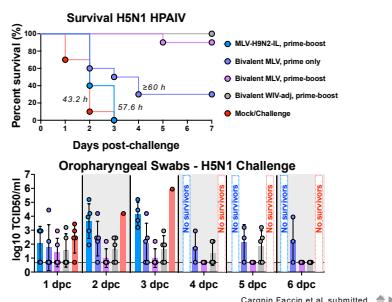
Monovalent and bivalent MLVs provide protection against drifted H9N2 challenge in contrast to bivalent WIV-adj vaccine



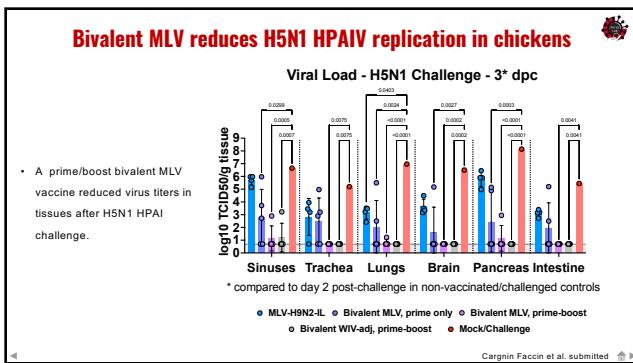
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Bivalent MLV provides protection against H5N1 HPAIV

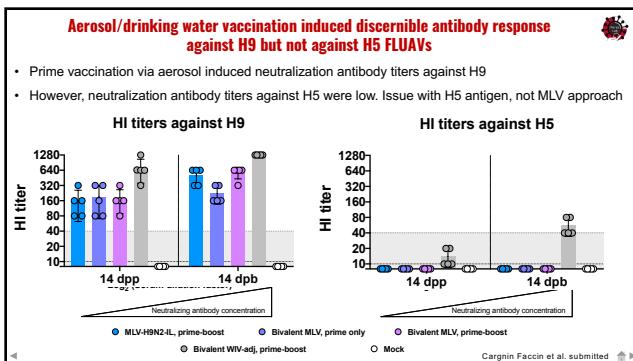
- Chickens were challenged with 50 ckLD₅₀ of H5N1 two weeks after boost (1.58×10^4 EID_{50/chicken})
- A bivalent vaccine administered through a prime-boost method resulted in a 90% protection rate and lower viral loads in comparison to other groups.



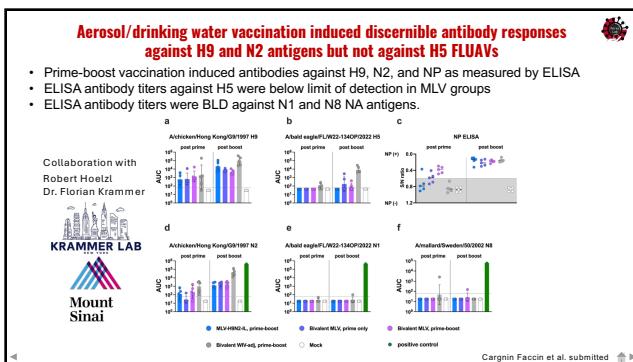
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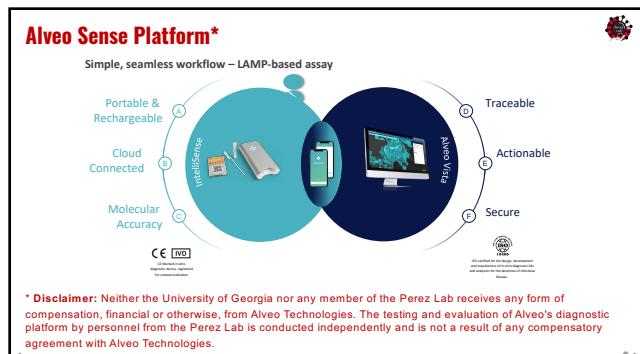
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Alveo Sense Poultry Avian Influenza Test Type A H5 H9

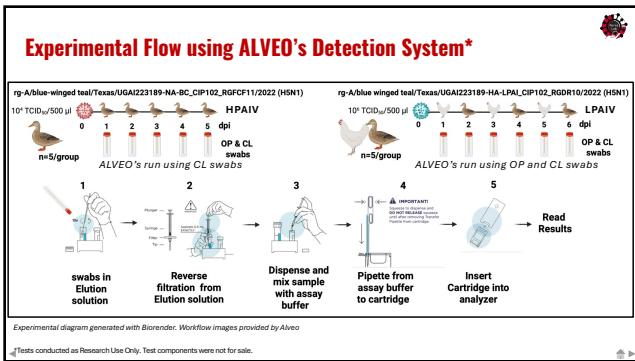
Test Design:

Limit of Detection:

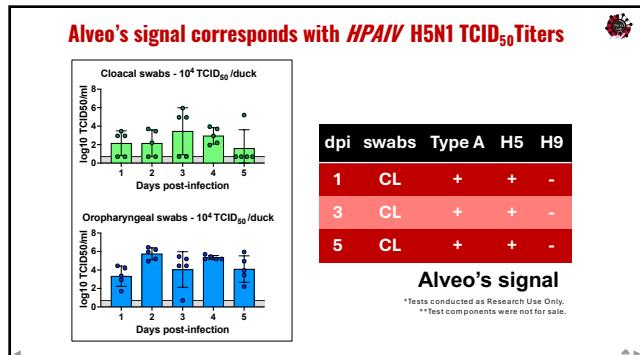
Target	Individual Assays	Sample	Matrix Type	Equivalent PCR Ct GD TaqMan PCR
Type A	2 unique regions of matrix gene for broad InflA detection	H5N2 (Am_h5n2gd)	Oropharyngeal (Pooling)	33.5
Subtype H5	Targeted cluster of H5NT: 2,3,4,6 (H5PA1) American non-specific lineage European non-specific lineage	H5N6 (2,3,4,6)	Oropharyngeal (Pooling)	33.0
Subtype H9	Detect clinically relevant H9 strains	H5N2 (G lineage)	Oropharyngeal (Pooling)	30.6
			Oropharyngeal (Pooling)	31.9
			Oropharyngeal (Pooling)	31.6
			Oropharyngeal (Pooling)	31.6

*** Disclaimer:** Neither the University of Georgia nor any member of the Perez Lab receives any form of compensation, financial or otherwise, from Alveo Technologies. The testing and evaluation of Alveo's diagnostic platform by personnel from the Perez Lab is conducted independently and is not a result of any compensatory agreement with Alveo Technologies.

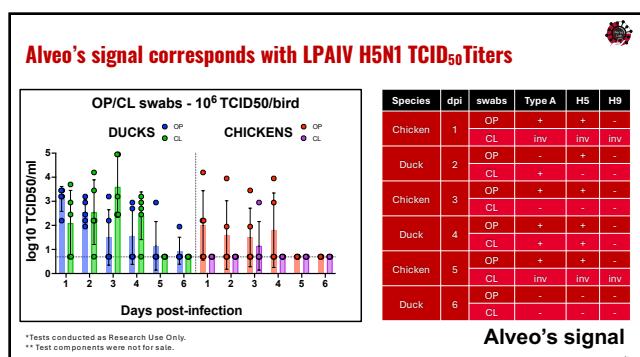
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Summary: MLVs to prevent and control avian influenza viruses

- MLVs offer an opportunity for mass vaccination – drinking water, aerosol, spray(?)
- MLVs are stable, immunogenic, do not transmit, and do not reassort (?)
- The inclusion of an immunomodulator (Flu-kin) enhanced the generation of neutralizing and non-neutralizing antibodies, suggesting a role in the host immune response.
- Aerosol vaccination results in limited MLV virus replication.
- Aerosol and drinking water vaccination elicits measurable serological antibody subtype specific responses against HA and NA. Opportunity for production of DIVA-compliant MLVs based on anti-NA responses? Anti-NP responses readily observed too.
- The bivalent MLV vaccine provides protection against homologous H5N1 HPAIV, despite negligible anti-H5 antibody responses.
- The bivalent H9 MLV vaccine provides protection against (homologous) and drifted H9N2 FLUAV challenges (contrasting WIV-adj. vaccine).



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Summary: Experimental use of ALVEO's portable diagnostic system

Intuitive, easy to operate, enables efficient result sharing.

Correctly identified 100% of expected results in ducks challenged with HPAIV-H5N1.

**LPAIV-H5N1 Challenged Ducks**

- Results aligned with viral load (based on TCID₅₀ measurements).
- OP positive for H5, CL positive for Type A at 2 DPI. These partial detections still represent actionable field results, as any positive signal would prompt confirmatory testing.

LPAIV-H5N1 Challenged Chickens

- Chickens exhibited different infection dynamics compared to ducks.
- OP pools performed as expected, confirmatory RT-PCR test underway on the OP pool from 5 DPI.
- CL pools produced two invalid results. The negative result at 3 DPI is likely attributable to low sample titer. The invalid result at 1 DPI may be due to a workflow deviation.
- 100% H9 exclusivity, no cross-reactivity.

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Avian Flu Rapid Diagnostics Moving Forward:

- Current lack of globally harmonized data on how well rapid tests (specifically LFDs) perform across different AIV clades, geographies, and sample types.
- (Relatively) cheap, portable, and fast diagnostic tests that can be used for immediate detection in poultry farms, markets, and outbreak zones are coming along; however, ...
- Tests must be able to detect not only the dominant H5N1 clades but also other evolving and reassorting strains endemic in poultry and that pose a pandemic threat.
- Governments and response teams need transparent, evidence-based data to support their procurement and deployment decisions, as many current kits are unproven against avian variants.
- RapidAV**, a Global Collaborative to Harmonize and Validate Rapid Diagnostic Tests for Avian Influenza proposed by Erik Karlsson Director, National Influenza Center of Cambodia and WHO H5 Reference Laboratory, Coordinator, WHO Global COVID-19 Referral Laboratory, Institut Pasteur du Cambodge aims to assemble a collaborative network with common metrics to overcome the scattered and isolated small-scale assessments currently being conducted by individual labs.



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